Regulation of Cerebral Cortical Size by Control of Cell Cycle Exit in Neural Precursors

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Transgenic mice expressing a stabilized β-catenin in neural precursors develop enlarged brains with increased cerebral cortical surface area and folds resembling sulci and gyri of higher mammals. Brains from transgenic animals have enlarged lateral ventricles lined with neuroepithelial precursor cells, reflecting an expansion of the precursor population. Compared with wild-type precursors, a greater proportion of transgenic precursors reenter the cell cycle after mitosis. These results show that β-catenin can function in the decision of precursors to proliferate or differentiate during mammalian neuronal development and suggest that β-catenin can regulate cerebral cortical size by controlling the generation of neural precursor cells.

A massive increase in the size of the cerebral cortex is thought to underlie the growth of intellectual capacity during mammalian evolution. The increased size of larger brains results primarily from a disproportionate expansion of the surface area of the layered sheet of neurons comprising the cerebral cortex (1–7), with the appearance of convolutions of the cortical surface (with crests known as gyri and intervening grooves called sulci) providing a means of increasing the total cortical area in a given skull volume. This horizontal expansion of the cerebral cortex is not accompanied by a comparable increase in cortical thickness; in fact, the 1000-fold increase in cortical surface area between human and mouse is only accompanied by an ∼two-fold increase in cortical thickness (8).

The cerebral cortex is organized into columnar functional units (9), and the expansion of the cerebral cortex appears to result from increases in the number of radial columns rather than from increases in individual column size (5, 10). These observations have led to the proposal that increases in the number of columns result from a corresponding increased number of progenitor cells (5). It has been suggested that minor changes in the relative production of progenitors and neurons could produce dramatic increases in cortical surface area (5, 11).

One protein that might regulate the production of neural precursors is β-catenin, an integral component of adherens junctions (12) that interacts with proteins of the T cell factor/lymphoid enhancer binding factor (TCF/LEF) family to transduce Wnt signals (13). Wnts (a family of secreted signaling proteins) are expressed in overlapping patterns in the developing mammalian brain, and numerous studies support the role of Wnt signaling in cell fate regulation during development (17). Inactivation of specific Wnts (18, 19), TCFC/LEF members (20), or β-catenin (21) results in specific developmental brain defects, and persistent activation of β-catenin has been implicated in a variety of human cancers (13), including some resembling neural precursors such as medulloblastoma (22). These findings raise the possibility that β-catenin influences cell number or cell fate decisions in the developing nervous system. β-catenin is widely expressed in many tissues (23). To examine more closely the expression patterns of β-catenin during mammalian

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Materials and Methods
Figs. S1 to S5
Movie S1

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neural development, in situ hybridization of β-catenin was performed on embryonic mouse brain sections. Strong hybridization was observed for β-catenin in neuroepithelial precursors in the ventricular zone across the period during which neurons were produced (Fig. 1A). Immunostaining with a monoclonal antibody indicates that, in neuroepithelial precursors, β-catenin protein is enriched at adherens junctions at the lumen of the ventricle, where it colocalizes in rings with F-actin, highlighted by rhodamine phalloidin (Fig. 1B).

To examine whether activating β-catenin signaling could regulate mammalian brain development, we generated transgenic mice overexpressing an NH₂-terminally truncated form of β-catenin fused at the COOH-terminus with green fluorescent protein (GFP) (Δ90β-catenin-GFP) in neuroepithelial precursors. NH₂-terminally truncated β-catenin no longer requires Wnt signaling for sustaining activity, because it lacks key phosphorylation sites for GSK3β that normally target it for destruction in the absence of Wnts (24).

This form of β-catenin is stabilized constitutively in vivo and remains able to bind E-cadherin and α-catenin and to activate transcription by binding with TCF/LEF cofactors (24, 25) (Fig. 2B) [see supplementary online material (SOM)]. The expression of Δ90β-catenin-GFP was driven by the enhancer element contained in the second intron of the nestin gene (Fig. 2C) (see SOM), which directs expression in central nervous system progenitor cells (26).

Transgenic embryos at embryonic day 15.5 (E15.5) have grossly enlarged brains, with a considerable increase in the surface area of the cerebral cortex, without a corresponding increase in cortical thickness (n = 10) (Fig. 3). Sections through the forebrain revealed that, in transgenic brains, the horizontal growth of the tissue is so extensive that the normally smooth cerebral cortex of the mouse forms undulating folds resembling the gyri and sulci of higher mammals (Fig. 3B) (27). Brains from E17.5 embryos showed similar enlargement and folding (fig. S1). In

![Fig. 1. Expression of β-catenin transcript and protein in neural precursors.](image1)

(A) β-catenin in situ hybridization in sections through developing mouse cerebral cortex. β-catenin is strongly expressed in the ventricular zone (VZ) precursor cells at all ages during which cortical neurons are generated. A weaker signal is present in the developing cortical plate. Bar, 200 μm. (B) Immunostaining through E14.5 mouse ventricular zone reveals β-catenin immunoreactivity (green) concentrated in rings at the lumenal surface. Staining of the same section with rhodamine phalloidin reveals F-actin (red), which colocalized with adherens junctions in a ringlike distribution at the lumenal surface. The merged view indicates that β-catenin colocalizes with phalloidin. Bar, 10 μm.

![Fig. 2. Transcriptional activation by β-catenin and expression and transgenic construct design.](image2)

(A) pTOPFLASH luciferase reporter assay in NT2 cells. NT-2 cells were transfected with pTOPFLASH, containing four consensus Lef-1/Tcf-1 binding sites, a minimal Fos promoter, and a luciferase reporter (43). Transfections were performed with and without cytomegalovirus (CMV)-Δ90β-catenin-GFP. CMV-LacZ was used to normalize for transfection efficiency. Twenty-four hours later, cells were lysed and protein extracts were assayed for luciferase. Fold inductions of luciferase activity represent the average of three experiments, with error bars representing one SEM. (B) Δ90β-catenin activates transcription in primary cortical cells. Primary cells from E17 cortex were transfected with the pTOPFLASH luciferase reporter construct and the expression vectors as indicated. Luciferase activity was assayed 48 hours after transfection. Fold inductions represent the average of six experiments, with error bars indicating one SEM. (C) Expression and transgenic constructs. Constructs removing the NH₂-terminal 90 amino acids of mouse β-catenin are fused either to EGFP or the kt3 epitope tag. For expression in transient transfection assays, β-catenin constructs are placed behind the CMV promoter. The nestin second intron coupled with the thymidine kinase minimal promoter are used to generate transgenic mice. The first intron from the rat insulin II gene is incorporated to enhance expression levels. The same β-catenin alleles were used in both in vitro and transgenic mice.
cresyl violet–stained sections, a densely stained layer of cells adjacent to the enlarged ventricular lumen morphologically resembled the proliferative zone of wild-type brains but was greatly expanded in surface area in the transgenic animals (E15.5, n = 10; E17.5, n = 6; E19.5, n = 2). Because we observed marked expansion of the cortical neuroepithelium, we focused our further studies on this population of cells at E15.5, an age midway through mouse cortical neurogenesis.

To determine the identity of the cells that may account for the expansion of the transgenic brains, we examined the expression of markers specific for neuroepithelial precursors and differentiating neurons. The basic helix-loop-helix transcription factors Hes5 and Hes1 are downstream effectors of the Notch signaling pathway and regulate neuronal differentiation. Hes5 is expressed specifically by neuroepithelial precursors, whereas Hes1 is highly expressed in precursors, with lower expression in more differentiated cortical plate neurons. In situ hybridization for Hes5 of comparable coronal sections through wild-type and transgenic brains suggests that the neural precursor population in transgenic animals is expanded (Fig. 4A). The expression of both Hes1 (Fig. S1) and Ki67 (Fig. 5), a protein expressed in all dividing cells, highlighted the ventricular zone and confirmed the findings seen with Hes5, providing further support that the precursor zone is expanded in transgenic animals. Finally, we used the thymidine analog BrdU to label dividing neural precursor cells by exposing embryos to BrdU for 30 min before killing them. Sections through wild-type and transgenic brains show that the same cells lining the ventricle also incorporate BrdU, confirming that the population of cells labeled with the precursor markers is composed of dividing cells (Fig. 3, E and F).

To investigate the spatial patterns of neuronal differentiation in transgenic animals, we examined the expression of three different markers of cortical neuron populations—Reelin (Reln), T-box brain gene 1 (Thrb1), and TuJ1. In wild-type mice at E15.5, Reln labels Cajal-Retzius neurons in the outermost rind of cells of the developing cortical plate (Fig. 4). Similarly, in the brains of transgenic animals, in situ hybridization for Reln expression showed strong labeling in its normal position at the margin of the cortical plate. In wild-type mice at E15.5, Thrb1 is normally expressed in neurons of the cortical preplate and subplate (Fig. 4). Similarly, in situ hybridization for Thrb1 in transgenic animals indicates that cortical cells outside the ventricular zone expressed Thrb1 (Fig. 4). The general pattern of Thrb1 staining resembled that of wild-type mice, with Thrb1–expressing cells situated in the region outside the progenitor zone in the developing cortical plate. However, much like those that express Reln, cells that express Thrb1 were somewhat more widely scattered throughout the developing cortical plate, as compared with cells with wild-type expression. In E15.5 wild-type animals, TuJ1 labels newly differentiated neurons outside the ventricular zone (Fig. 4). In transgenic mice, TuJ1 immunoreactivity also labeled the layer of cells outside the ventricular zone, supporting the idea that postmitotic neurons remain localized outside the ventricular zone in transgenic animals. Despite the massive expansion of cortical surface area, transgenic precursors appear to differentiate into young neurons in an approximately normal spatial pattern. Taken together, these expression studies suggest that over-activating β-catenin does not disrupt the normal developmental sequence of neuronal differentiation, and the horizontal expansion of the cortical plate is a result of an increased number of proliferative precursor cells.

Enlargement of the precursor pool in transgenic brains can result from increased mitotic rates, decreased cell death, changes in cell fate choice (whether to differentiate or to proliferate), or any combination of these factors. To examine whether the horizontal expansion of the progenitor pool in transgenic animals results from increased mitotic rates, we counted the proportion of precursor cells that could be labeled by a 30-min pulse of BrdU. To quantify the fraction of cells that are cycling, we obtained a labeling index (LI) by counting the percentage of cortical progenitor cells that were labeled by a single pulse of BrdU.
BrdU. Progenitor cells were identified by Ki67 immunoreactivity (30, 31). Because in mammalian cells the length of S phase remains relatively constant while the length of G1 regulates proliferation (32), this LI provides an estimation of cell cycle length. If the cell cycle is shortened, the relative fraction of cells labeled by a brief BrdU pulse will increase. Examination of random fields chosen from six brains (three wild-type and three transgenic brains) suggests that the transgenic neural precursors did not divide significantly faster than did normal wild-type precursors [F(6,36) = 0.970, P = 0.471] (Fig. 5A).

Programmed cell death (apoptosis) occurs during normal development of the central nervous system (33), and decreased programmed cell death may be one mechanism underlying the increased brain size of transgenic animals. Apoptotic cell death was examined using TUNEL staining in wild-type and transgenic brains. TUNEL+ cells were confirmed by verifying condensed nuclei labeled with the DNA binding dye Hoechst 33342. Counts of total numbers of labeled cells revealed that cell death in transgenic brains was not substantially less than found in wild type (Fig. 5B); in fact, there appeared to be greater than twofold increased rates of apoptosis in transgenic brains [F(6,11) = 26.00, P = 0.0002]. Taken together, the BrdU-labeling studies and TUNEL studies suggest that the progenitor cell population expansion cannot be explained by a simple mitogenic effect of β-catenin or by decreased apoptotic cell death.

Progenitor divisions that give rise to additional progenitors can expand the progenitor pool exponentially. Consequently, small alterations in the fraction of cell divisions that expand the progenitor pool can result in large changes in the final size of the brain (3, 34). To examine whether the increase in the progenitor pool results from a shift in the fraction of progenitors that choose to remain progenitors instead of differentiating, we examined cell cycle exit and re-entry by examining the fraction of cells dividing after pulse labeling with BrdU 24 hours earlier. We identified cells that had left the cell cycle as BrdU+ and Ki67−, and we identified cells that remained in the cell cycle as BrdU+ and Ki67+. At E15.5, we found an ~twofold increase in the proportion of transgenic precursors that re-enter the cell cycle when compared with wild-type neural precursors [F(4, 15) = 11.00, P = 0.0009] (Fig. 5C).

Together, these studies suggest that β-catenin activation functions in neural precursors to influence the decision to re-enter the cell cycle instead of differentiating.

Our results support recent findings suggesting that epithelial architecture and adherens junctions regulate growth control and cell proliferation (35). Because β-catenin is an integral component of adherens junctions (12), disruptions of adherens junctions may cause misregu-
loration and accumulation of cytoplasmic β-cate
nin. Our findings that β-catenin signaling can re
duce the decisions of neural precursors to re
enter or exit the cell cycle lend support to the pos
sibility that β-catenin signaling may mediate the loss of growth control when adherens junc
tions are disrupted.

It has been hypothesized that mutations in reg
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precursors to divide or differentiate can underlie the expansion of the precursor popu
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**References and Notes**


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**Nonresonant Multiple Spin Echoes**

Thilo M. Brill,* Seungoh Ryu, Richard Gaylor, Jacques Jundt, Douglas D. Griffin, Yi-Qiao Song, Pabitra N. Sen, Martin D. Hürlimann

Nonresonant manipulation of nuclear spins can probe large volumes of sample
situated in inhomogeneous fields outside a magnet, a geometry suitable for
mobile sensors for the inspection of roads, buildings, and geological formations.
However, the interference by Earth’s magnetic field causes rapid decay of the signal within a few milliseconds for protons and is detrimental to this method.
Here we describe a technique to suppress the effects of Earth’s field by using
adiabatic rotations and sudden switching of the applied fields. We observed hundreds of spin echo signals lasting for more than 600 milliseconds and accurately measured the relaxation times of a liquid sample.

Conventional nuclear magnetic resonance (NMR) experiments are almost always car
ried out by manipulating nuclear spins using radio frequency (rf) pulses at the spin Larmor frequency \(\omega = \gamma B\), where \(\gamma\) is the gyromagnetic ratio and \(B\) is the magnitude of the magnetic field. Such resonant NMR experi
ments allow the imaging of spins in materials and the characterization of spin interactions, enab
ling applications extending to materials such as soft condensed matter (1), plants (2), food products (3), cement and concrete (4), and geological materials (5, 6). The field applications are the motivation for several recent developments in ex situ NMR (7–10), where a mobile NMR detector is used to examine the sample outside the NMR mag
net. However, as a result of the geometry of such mobile tools, the applied magnetic fields exhibit large inhomogeneities, and all reso
nant techniques will result in small sensitive volumes where the resonance condition is satis
fied. Composite (11) and adiabatic (12) pulses may be used to expand the excitation bandwidth to a limited extent at the expense of higher irradiation power.

Alternatively, spins can be manipulated...